(capillary columns). A capillary column has a maximum internal diameter (Ø) of 100 µm.

**Mobile phases**

Usually the mobile phase is carbon-dioxide which may contain a polar modifier such as methanol, 2-propanol or acetonitrile. The composition, pressure (density), temperature and flow rate of the prescribed mobile phase may either be constant throughout the whole chromatographic procedure (isocratic, isodense, isothermic elution) or may vary according to a defined programme (gradient elution of the modifier, pressure (density), temperature or flow rate).

**Detectors**

Ultraviolet/visible (UV/Vis) spectrophotometers and flame ionisation detectors are the most commonly employed detectors. Light scattering detectors, infrared absorption spectrophotometers, thermal conductivity detectors or other special detectors may be used.

**METHOD**

Prepare the test solution(s) and the reference solution(s) as prescribed. The solutions must be free from solid particles.

Criteria for assessing the suitability of the system are described in the chapter on Chromatographic separation techniques (2.2.46). The extent to which adjustments of parameters of the chromatographic system can be made to satisfy the criteria of system suitability are also given in this chapter.

**2.2.46. CHROMATOGRAPHIC SEPARATION TECHNIQUES**

Chromatographic separation techniques are multi-stage separation methods in which the components of a sample are distributed between 2 phases, one of which is stationary, while the other is mobile. The stationary phase may be a solid or a liquid supported on a solid or a gel. The stationary phase may be packed in a column, spread as a layer, or distributed as a film, etc. The mobile phase may be gaseous or liquid or supercritical fluid. The separation may be based on adsorption, mass distribution (partition), ion exchange, etc., or may be based on differences in the physico-chemical properties of the molecules such as size, mass, volume, etc.

This chapter contains definitions and calculations of common parameters and generally applicable requirements for system suitability. Principles of separation, apparatus and methods are given in the following general methods:

- paper chromatography (2.2.26),
- thin-layer chromatography (2.2.27),
- gas chromatography (2.2.28),
- liquid chromatography (2.2.29),
- size-exclusion chromatography (2.2.30),
- supercritical fluid chromatography (2.2.45).

**DEFINITIONS**

The following definitions have been used to calculate the limits in monographs.

**EXAMPLE**

With some equipment, certain parameters, such as the signal-to-noise ratio, can be calculated using software provided by the manufacturer. It is the responsibility of the user to ensure that the calculation methods used in the software are compatible with the requirements of the European Pharmacopoeia. If not, the necessary corrections must be made.

**Chromatogram**

A chromatogram is a graphical or other representation of detector response, effluent concentration or other quantity used as a measure of effluent concentration, versus time, volume or distance. Idealised chromatograms are represented as a sequence of gaussian peaks on a baseline.

**RETENTION DATA**

**Retention time and retention volume**

Retention measurements in elution chromatography may be given as the retention time (t<sub>R</sub>) directly defined by the position of the maximum of the peak in the chromatogram. From the retention time, the retention volume (V<sub>R</sub>) may be calculated.

\[
V_R = v \times t_R
\]

\( t_R \) = retention time or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to the component,

\( v \) = flow rate of the mobile phase.

**Mass distribution ratio**

The mass distribution ratio (D<sub>m</sub>) (also known as the capacity factor k’ or retention factor k) is defined as:

\[
D_m = \frac{\text{amount of solute in stationary phase}}{\text{amount of solute in mobile phase}} = \frac{K_C}{V_M} \frac{V_S}{V_M}
\]

\( K_C \) = equilibrium distribution coefficient (also known as distribution constant),

\( V_M \) = volume of the mobile phase,

\( V_S \) = volume of the stationary phase.

The mass distribution ratio of a component may be determined from the chromatogram using the expression:

\[
D_m = \frac{t_R - t_M}{t_M}
\]

\( t_R \) = retention time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to the component,

\( t_M \) = hold-up time (or volume): time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the unretained component.

**Distribution coefficient**

The elution characteristics of a component in a particular column, in size-exclusion chromatography, may be given by
The distribution coefficient \( K_o \) which is calculated from the expression:

\[
K_o = \frac{t_{R} - t_{u}}{t_{f} - t_{u}}
\]

- \( t_{R} \) = retention time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to the component,
- \( t_{u} \) = hold-up time (or volume): time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to an unretained component,
- \( t_{f} \) = retention time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to a component which has full access to the pores of the stationary phase.

Retardation factor

The retardation factor \( R_F \) (also known as retention factor \( R_f \)), used in planar chromatography, is the ratio of the distance from the point of application to the centre of the spot and the distance travelled by the solvent front from the point of application.

\[
R_F = \frac{b}{a}
\]

- \( b \) = migration distance of the analyte,
- \( a \) = migration distance of the solvent front.

CHROMATOGRAPHIC DATA

The peak may be defined by the peak area \( (A) \) or the peak height \( (h) \) and the peak width at half-height \( (w_{1/2}) \) or the peak height \( (h) \) and the peak width between the points of inflection \( (w_i) \). In gaussian peaks (Figure 2.2.46.-1) there is the relationship:

\[
w_h = 1.18w_i
\]

Symmetry factor

The symmetry factor \( A_s \) (or tailing factor) of a peak (Figure 2.2.46.-2) is calculated from the expression:

\[
A_s = \frac{w_{0.05}}{2d}
\]

- \( w_{0.05} \) = width of the peak at one-twentieth of the peak height,
- \( d \) = distance between the perpendicular dropped from the peak maximum and the leading edge of the peak at one-twentieth of the peak height.

A value of 1.0 signifies complete (ideal) symmetry.

Column performance and apparent number of theoretical plates

The column performance (apparent efficiency) may be calculated from data obtained under either isothermal, isocratic or isodense conditions, depending on the technique, as the apparent number of theoretical plates \( (N) \) from the following expression, where the values of \( t_{R} \) and \( w_{1/2} \) have to be expressed in the same units (time, volume or distance).

\[
N = 5.54 \left( \frac{1R}{w_{1/2}} \right)^2
\]

- \( t_{R} \) = retention time (or volume) or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to the component,
- \( w_{1/2} \) = width of the peak at half-height.

The apparent number of theoretical plates varies with the component as well as with the column and the retention time.

SEPARATION DATA

Resolution

The resolution \( (R_s) \) between peaks of 2 components may be calculated from the expression:

\[
R_s = \frac{1.18(t_{R2} - t_{R1})}{w_{h1} + w_{h2}}
\]

- \( t_{R1} \) and \( t_{R2} \) = retention times or distances along the baseline from the point of injection to the perpendiculars dropped from the maxima of 2 adjacent peaks,
- \( w_{h1} \) and \( w_{h2} \) = peak widths at half-height.

A resolution of greater than 1.5 corresponds to baseline separation.

The expression given above may not be applicable if the peaks are not baseline separated.
In quantitative planar chromatography, the migration distances are used instead of retention times and the resolution may be calculated using the expression:

\[ R_s = \frac{1.18 \alpha (R_{F2} - R_{F1})}{w_{h1} + w_{h2}} \]

Where:
- \( R_{F1} \) and \( R_{F2} \) are ratios of the distances from the point of application to the centres of the spots and the distance travelled by the solvent front from the point of application (retardation factor),
- \( w_{h1} \) and \( w_{h2} \) are peak widths at half-height,
- \( \alpha \) is the migration distance of the solvent front.

**Peak-to-valley ratio**

The peak-to-valley ratio \((p/v)\) may be employed as a system suitability requirement for related substances when baseline separation between 2 peaks is not reached (Figure 2.2.46.-3).

\[ p/v = \frac{H_p}{H_v} \]

Where:
- \( H_p \) is the height above the extrapolated baseline of the minor peak,
- \( H_v \) is the height above the extrapolated baseline at the lowest point of the curve separating the minor and major peaks.

**Relative retention**

The relative retention \((r)\) is calculated as an estimate from the expression:

\[ r = \frac{t_{R2} - t_M}{t_{R1} - t_M} \]

Where:
- \( t_{R2} \) is the retention time of the peak of interest,
- \( t_{R1} \) is the retention time of the reference peak (usually the peak corresponding to the substance to be examined),
- \( t_M \) is the hold-up time: time or distance along the baseline from the point of injection to the perpendicular dropped from the maximum of the peak corresponding to an unretained component.

The unadjusted relative retention \((r_G)\) is calculated from the expression:

\[ r_G = \frac{t_{R2}}{t_{R1}} \]

Unless otherwise indicated, values for relative retention stated in monographs correspond to unadjusted relative retention.

In planar chromatography, the retardation factors \( R_{F2} \) and \( R_{F1} \) are used instead of \( t_{R2} \) and \( t_{R1} \).

**PRECISION OF QUANTIFICATION**

**Signal-to-noise ratio**

The signal-to-noise ratio \((S/N)\) influences the precision of quantification and is calculated from the equation:

\[ S/N = \frac{2H}{h} \]

Where:
- \( H \) is the height of the peak (Figure 2.2.46.-4) corresponding to the component concerned, in the chromatogram obtained with the prescribed reference solution, measured from the maximum of the peak to the extrapolated baseline of the signal observed over a distance equal to 20 times the width at half-height,
- \( h \) is the range of the background noise in a chromatogram obtained after injection or application of a blank, observed over a distance equal to 20 times the width at half-height of the peak in the chromatogram obtained with the prescribed reference solution and, if possible, situated equally around the place where this peak would be found.

**Repeatability**

The repeatability of response is expressed as an estimated percentage relative standard deviation \((RSD_p)\) of a consecutive series of measurement of injections or
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The maximal permitted relative standard deviation (RSD\(_{\text{max}}\)) is calculated for a series of injections of the reference solution for defined limits using the following expression:

\[
RSD_{\text{max}} = \frac{KB\sqrt{n}}{t_{95\%} n-1}
\]

- \(K\) = constant (0.349), obtained from the expression \(K = \frac{0.674}{\sqrt{n}}\), in which \(\frac{0.674}{\sqrt{n}}\) represents the required RSD after 6 injections for \(B = 1.0\),
- \(B\) = upper limit given in the definition of the individual monograph minus 100 per cent,
- \(n\) = number of replicate injections of the reference solution (3 \(\leq n \leq 6\)),
- \(t_{95\%,n-1}\) = Student’s \(t\) at the 90 per cent probability level (double sided) with \(n-1\) degrees of freedom.

**SYSTEM SUITABILITY**

The system suitability tests represent an integral part of the method and are used to ensure adequate performance of the chromatographic system. Apparent efficiency, mass distribution ratio, resolution, relative retention and the symmetry factor are the parameters which are usually employed in assessing the performance of the column. Factors which may affect the chromatographic behaviour include the composition, ionic strength, temperature and apparent pH of the mobile phase, flow rate, column length, temperature and pressure, and stationary phase characteristics including porosity, particle size, type of particles, specific surface area and, in the case of reverse-phase supports, the extent of chemical modification (as expressed by end-capping, carbon loading etc.).

The various components of the equipment employed must be qualified and be capable of achieving the precision required to conduct the test or assay.

The following requirements are to be fulfilled unless otherwise stated in the monograph:

- The symmetry factor of the principal peak is to be between 0.8 and 1.5 unless otherwise stated in the monograph. This requirement has general applicability to tests or assays described in the monographs.
- Maximal permitted relative standard deviation for replicate injections of the prescribed reference solution do not exceed the values given in Table 2.2.46.-1. This requirement is applicable to assays for content only and does not apply to the test for related substances.
- The limit of detection of the peak (corresponding to a signal-to-noise ratio of 3) is below the disregard limit of the test for related substances.
- The limit of quantitation of the peak (corresponding to a signal-to-noise ratio of 10) is equal to or less than the disregard limit of the test for related substances.

**ADJUSTMENT OF CHROMATOGRAPHIC CONDITIONS**

The extent to which the various parameters of a chromatographic test may be adjusted to satisfy the system suitability criteria without fundamentally modifying the methods is listed below for information. The chromatographic conditions described have been validated during the elaboration of the monograph. The system suitability tests are included to ensure the separation required for satisfactory performance of the test or assay. Nonetheless, since the stationary phases are described in a general way and there is such a variety available commercially, with differences in chromatographic behaviour, some adjustments of the chromatographic conditions may be necessary to achieve the prescribed system suitability requirements. With reverse-phase liquid chromatographic methods, in particular, adjustment of the various parameters will not always result in satisfactory chromatography. In that case, it may be necessary to replace the column with another of the same type (e.g. octadecylsilil silica gel) which exhibits the desired chromatographic behaviour.

For critical parameters the adjustments are defined clearly in the monograph to ensure the system suitability. Multiple adjustments which may have a cumulative effect in the performance of the system are to be avoided.

**Thin-layer chromatography and paper chromatography**

**Composition of the mobile phase:** the amount of the minor solvent component may be adjusted by ± 30 per cent relative or ± 2 per cent absolute, whichever is the larger; for a minor component at 10 per cent of the mobile phase, a 30 per cent relative adjustment allows a range of 7-13 per cent whereas a 2 per cent absolute adjustment allows a range of 8-12 per cent, the relative value being therefore the larger; for a minor component at 5 per cent of the mobile phase, a 30 per cent relative adjustment allows a range of 3.5-6.5 per cent whereas a 2 per cent absolute adjustment allows a range of 3-7 per cent, the absolute value being in this case the larger. No other component is altered by more than 10 per cent absolute.

**pH of the aqueous component of the mobile phase:** ± 0.2 pH, unless otherwise stated in the monograph, or ± 1.0 pH when neutral substances are to be examined.

**Concentration of salts in the buffer component of a mobile phase:** ± 10 per cent.

**Application volume:** 10-20 per cent of the prescribed volume if using fine particle size plates (2-10 µm).

**Migration distance of the solvent front** is to be not less than 50 mm or 30 mm on high-performance plates.

**Liquid chromatography**

**Composition of the mobile phase:** the amount of the minor solvent component may be adjusted by ± 30 per cent relative or ± 2 per cent absolute, whichever is the larger (see example above). No other component is altered by more than 10 per cent absolute.
pH of the aqueous component of the mobile phase: ± 0.2 pH, unless otherwise stated in the monograph, or ± 1.0 pH when neutral substances are to be examined.

Concentration of salts in the buffer component of the mobile phase: ± 10 per cent.

Detector wavelength: no adjustment permitted.

Stationary phase:
- column length: ± 70 per cent,
- column internal diameter: ± 25 per cent,
- particle size: maximal reduction of 50 per cent, no increase permitted.

Flow rate: ± 50 per cent. When in a monograph the retention time of the principle peak is indicated, the flow rate has to be adjusted if the column internal diameter has been changed. No decrease of flow rate is permitted if the monograph uses an apparent number of theoretical plates in the qualification section.

Temperature: ± 10 per cent, to a maximum of 60 °C.

Injection volume: may be decreased, provided detection and repeatability of the peak(s) to be determined are satisfactory.

Gradient elution: the configuration of the equipment employed may significantly alter the resolution, retention time and relative retentions described in the method. Should this occur, it may be due to excessive dwell volume which is the volume between the point at which the 2 eluants meet and the top of the column.

Gas chromatography

Stationary phase:
- column length: ± 70 per cent,
- column internal diameter: ± 50 per cent,
- particle size: maximal reduction of 50 per cent, no increase permitted,
- film thickness: −50 per cent to + 100 per cent.

Flow rate: ± 50 per cent.

Temperature: ± 10 per cent.

Injection volume: may be decreased, provided detection and repeatability are satisfactory.

Supercritical fluid chromatography

Composition of the mobile phase: for packed columns, the amount of the minor solvent component may be adjusted by ± 30 per cent relative or ± 2 per cent absolute, whichever is the larger. No adjustment is permitted for a capillary column system.

Detector wavelength: no adjustment permitted.

Stationary phase:
- column length: ± 70 per cent,
- column internal diameter:
  - ± 25 per cent (packed columns),
  - ± 50 per cent (capillary columns),
- particle size: maximal reduction of 50 per cent, no increase permitted (packed columns).

Flow rate: ± 50 per cent.

Temperature: ± 10 per cent.

Injection volume: may be decreased, provided detection and repeatability are satisfactory.

Quantification

- Detector response. The detector sensitivity is the signal output per unit concentration or unit mass of a substance in the mobile phase entering the detector. The relative detector response factor, commonly referred to as the response factor, expresses the sensitivity of a detector relative to a standard substance. The correction factor is the reciprocal of the response factor.

- External standard method. The concentration of the component(s) to be analysed is determined by comparing the response(s) (peak(s)) obtained with the test solution to the response(s) (peak(s)) obtained with a reference solution.

- Internal standard method. Equal amounts of a component that is resolved from the substance to be examined (the internal standard) is introduced into the test solution and a reference solution. The internal standard should not react with the substance to be examined; it must be stable and must not contain impurities with a retention time similar to that of the substance to be examined. The concentration of the substance to be examined is determined by comparing the ratio of the peak areas or peak heights due to the substance to be examined and the internal standard in the test solution with the ratio of the peak areas or peak heights due to the substance to be examined and the internal standard in the reference solution.

- Normalisation procedure. The percentage content of one or more components of the substance to be examined is calculated by determining the area of the peak or peaks as a percentage of the total area of all the peaks, excluding those due to solvents or any added reagents and those below the disregard limit.

- Calibration procedure. The relationship between the measured or evaluated signal (y) and the amount (concentration, mass, etc.) of substance (x) is determined and the calibration function is calculated. The analytical results are calculated from the measured signal or evaluated signal of the analyte by means of the inverse function.

For assays and for quantitative determination of components in the external standard method, the internal standard method or the calibration procedure may be described in the monograph, and the normalisation procedure is not normally applied. In tests for related substances, either the external standard method with a single reference solution or the normalisation procedure is generally applied. However, with both the normalisation procedure or the external standard method, when a dilution of the test solution is used for comparison, the responses of the related substances are similar to the substance itself (response factor of 0.8 to 1.2), otherwise correction factors are included in the text.

When the related substances test prescribes the summation of impurities or there is quantitative determination of an impurity, it is important to choose an appropriate threshold setting and appropriate conditions for the integration of the peak areas. In such tests the disregard limit, e.g. the areas of peaks whose areas are below the limit are not taken into account, is generally 0.05 per cent. Thus, the threshold setting of the data collection system corresponds to, at least, half of the disregard limit. Integration of the peak areas of the impurities, which are not completely separated from the main peak, are preferably performed by valley-to-valley extrapolation (tangential skim). Peaks due to the solvent(s) used to dissolve the sample are also to be disregarded.